

Online ISSN:2583-0376

http://jpps.ukaazpublications.com

DOI: http://dx.doi.org/10.54085/jpps.2023.3.3.1

Journal of Phytonanotechnology and Pharmaceutical Sciences

Review Article : Open Access

Diabetic nephropathy: An outline on molecular mechanism and protective pathways of phytoconstituents

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Article Info	Abstract		
Article history	Diabetic nephropathy is a debilitating complication of diabetes mellitus characterized by progressive		
Received 10 July 2023	kidney damage. This review provides a comprehensive overview of the molecular pathways underlying its		
Revised 12 August 2023	pathogenesis and explores the protective mechanisms offered by various phytobioactives. The pathogenetic		
Accepted 13 August 2023	mechanisms of diabetic nephropathy are multifaceted, encompassing the activation of key pathway		
Published Online 30 September 2023	such as the rennin angiotensin-aldosterone system (RAAS), the polyol pathway, protein kinase C (PKC)		
	pathway, hexosamine pathway, and the formation of advanced glycation end products (AGEs).		
Keywords	Understanding these pathways is crucial for developing effective therapeutic strategies to combat diabetic		
Diabetes	nephropathy. In parallel, this review sheds light on the role of phytobioactives, including alkaloid		
Nephropathy	flavonoids, tannins, and stilbenes, in mitigating the progression of diabetic nephropathy. These natural		
Inflammation	compounds exhibit a spectrum of protective properties, including antioxidative and anti-inflammatory		
Phytochemicals	effects. Their ability to modulate the molecular pathways implicated in diabetic nephropathy offers		
Flavonoids	promising avenues for novel therapeutic interventions. This review of the pathogenetic mechanisms and		
	the protective potential of phytobioactives underscore the importance of considering natural compounds		
	as adjunctive therapies in managing diabetic nephropathy, potentially improving patient outcomes and quality of life.		

1. Introduction

Due to its rising frequency and incidence, diabetes mellitus (DM) poses a major risk to the health of all people. In 2019, there were 463 million persons worldwide who had DM. By 2045, the International Diabetes Federation (IDF) predicts that this number would have surpassed 700 million (IDF, 2023; Kumaraswamy et al., 2022). A chronic metabolic condition known as diabetes mellitus (DM) causes higher than normal blood sugar levels (hyperglycemia) as a result of impaired insulin secretion, cellular resistance to insulin, or both (Punthakee et al., 2018). There are two main varieties of diabetes: type 1, in which the pancreas cannot generate insulin, and type 2, in which the body cannot effectively use the insulin that is produced (Latha and Vijaykumar, 2019). Microvascular consequences caused by DM include diabetic nephropathy (DN), diabetic retinopathy, and diabetic neuropathy in addition to macrovascular issues such stroke, cardiovascular disease (CVDs), and peripheral vascular disease (Adapa and Sarangi, 2015; Okur et al., 2017).

Diabetic nephropathy, also known as diabetic kidney disease, is a persistent renal ailment that has the potential to impact individuals diagnosed with either type 1 or type 2 diabetes. About 40% of all

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Copyright © 2023 Ukaaz Publications. All rights reserved. Email: ukaaz@yahoo.com; Website: www.ukaazpublications.com cases of diabetes are affected by DN, and projections indicate that by 2023, there will be 382 million cases of diabetes worldwide, or 8-10% of the world's population (Lim, 2014). Diabetic nephropathy (DN) is characterized by a plethora of clinical manifestations, including heightened arterial blood pressure, a compromised glomerular filtration rate (GFR), the presence of diabetic glomerular lesions, and escalated urine albumin excretion levels surpassing the threshold of 300 mg/day (Gheith et al., 2016). High blood sugar damages renal blood vessels, which leads to kidney dysfunction and is the underlying cause of DN (Gajjala et al., 2015). There are five stages of kidney degeneration associated with DN, and stage 4 is typically when symptoms first appear. It is advised that diabetics have annual renal problems screenings because symptoms do not manifest until later stages. The swelling of ankles, legs, and hands due to water retention, blood in the urine, exhaustion from low oxygen levels in the blood, and nausea are the defining characteristics of the fourth stage of DN. If left unaddressed, this can progress to end-stage renal disease (ESRD), which is the fifth and final stage, where dialysis or kidney transplantation are the only viable treatment options as the kidneys can no longer function to meet daily needs. The risk factors associated with developing DN include hypertension dyslipidemia, smoking and poor glycemic control. A person's genetic makeup also has a significant impact on developing DN because those with a family history of the condition are more likely to do so (Ahmad et al., 2013). According to current reports, DN accounts for 30 - 40% of ESRD cases in the US and is one of the main causes of ESRD (Ghaderian et al., 2015).

2. Pathogenesis of DN

The etiology of diabetic nephropathy (DN) is multifactorial. The onset of hyperglycemia is a pivotal event that triggers structural and functional alterations in the renal system, such as glomerular hyperfiltration, hypertrophy of glomerular and tubular epithelium, and microalbuminuria. Eventually, thickening of the glomerular basement membrane, accumulation of mesangial matrix, overt proteinuria, glomerulosclerosis, and end-stage renal disease (ESRD) ensue. There are number of pathways involved in pathogenesis of DN (Figure 1).

2.1 Protein kinase C pathways

The pathogenesis of diabetic nephropathy (DN) has been associated with elevated flux of glucose through the hexosamine pathways, leading to an increase in TGF- β expression (Kolm-Litty *et al.*, 1998). This occurs due to the glycosylation of transcription factor Sp1 by N-acetylglucosamine, causing fructose-6-phosphate from glycolysis to convert to glucosamine-6-phosphate, thereby promoting TGF- β transcription. The hexosamine pathway also upregulates the expression of upstream stimulatory factors (USFs), which activate the TGF 1 promoter (Weigert et al., 2004). Furthermore, intracellular glucose accumulation increases the de novo production of diacylglycerol (DAG) from glycolytic intermediates such as dihydroxyacetone phosphate (Haneda et al., 2003). Consequently, the activation of protein kinase C isoforms by DAG and inhibition of PKC, the main isoform generated by hyperglycemia in the kidney, reduces DN. Activation of PKC may also stimulate MAPKs, including Erk 1,2 and p38 MAPK, which have been implicated in DN signaling processes (Haneda et al., 2003). Additionally, ROS activation of MAPKs may lead to cross-talk between the various pathways (Heilig et al., 2001). Recent findings have demonstrated the significance of PKC in the development of various DN alterations, as albuminuria was found to be absent in diabetic PKC- α deficient mice, although PKC- α deficiency had no effect on glomerular hypertrophy or the up-regulation of TGF-B (Menne et al., 2004).

2.2 Advance glycation end products

In chronic hyperglycemia, the extra glucose reacts with tissue proteins or free amino acids. DN is created as a result of this glycosylation. Initial products of this process are reversible early glycosylation products, and later, AGE. Because of the rise in AGEs, the glomerular epithelial cells' matrix proteins accumulate, collagenase activity declines, and the tight connection between the cells becomes defective (Singh *et al.*, 1998).

2.3 Oxidative stress

The metabolic activity of the nephron generates a significant amount of reactive oxygen species, which are counterbalanced by numerous antioxidant enzymes and free radical scavenging systems. These reactive oxygen species are responsible for harmful biological effects, such as protein oxidation, renal vasoconstriction, and DNA damage. Unfortunately, hyperglycemia exacerbates the production of reactive oxygen species, primarily within the mitochondria (Nishikawa *et al.*, 2007). Reactive oxygen species also contribute to the adverse pathways that glucose may take when hyperglycemia occurs, such as PKC activation and advanced glycation end product formation (Kiritoshi *et al.*, 2003; Vasavada *et al.*, 2005). Even prior to the onset of diabetes, hyperglycemia induces oxidative stress. In patients with more severe nephropathy (*i.e.*, proteinuria versus microalbuminuria), there are higher levels of DNA damage indicators triggered by reactive oxygen species. Additionally, biopsies of human kidney specimens from individuals with diabetes have revealed glyco-oxidation products (*i.e.*, combined products of glycation and protein oxidation) and lipoxidation in the mesangial matrix and glomeruli, which are less prevalent in specimens from non-diabetic individuals (Suzuki *et al.*, 1999).

2.4 Polyol pathways

The process of converting glucose to sorbitol through aldose reductase and subsequently to fructose by sorbitol dehydrogenase is observed within the polyol pathway. An increase in glucose uptake by the cell leads to a greater influx of glucose into the polyol pathway. The reduction of glucose to sorbitol necessitates the depletion of NADPH in cells, which is a vital substrate for the regeneration of glutathione. This, in turn, exacerbates intracellular oxidative stress. It is crucial to note that three deoxyglucone, an intermediate substance, serves as a precursor for AGEs (Haneda *et al.*, 2003; Bernobich *et al.*, 2004).

2.5 Renin angiotensin aldosterone system (RAAS)

Diabetic nephropathy is a serious complication of diabetes mellitus that arises due to a complex cascade of physiological processes. It begins with the activation of the renin angiotensin aldosterone system (RAAS). In response to various factors such as elevated blood sugar levels, the body increases the release of renin, an enzyme produced by the kidneys. Renin sets off a chain reaction, leading to the production of Angiotensin II, a potent vasoconstrictor. As Angiotensin II levels rise, it causes the blood vessels in the kidneys to constrict, resulting in increased pressure within the glomeruli, the tiny filtration units in the kidneys. This heightened pressure, over time, damages the delicate kidney cells, particularly those responsible for filtration. As a consequence of this damage, these cells become less effective at retaining essential proteins, such as albumin, within the bloodstream. The loss of albumin into the urine, known as albuminuria, is a hallmark sign of diabetic nephropathy. Over time, if left uncontrolled, this condition can progress to more severe kidney dysfunction, ultimately leading to end-stage renal disease (Rahimi, 2016; Sato et al., 2003).

3. Phytochemicals

A plant-based diet's many health benefits are attributable to phytochemicals, which are organic compounds found in plants (Liu, 2013). Phytochemicals are classified into numerous classes according to their chemical composition, each of which has unique properties and potential health advantages. Phytochemical subgroups such as carotenoids, flavonoids, phenolic acids, alkaloids, and terpenoids are frequently found (Koche *et al.*, 2016). It has been shown that many phytochemicals have pharmacological action, which implies that they may interact with the body to produce benefits for the cardiovascular system, nervous system, anticancer, antiviral, and anti-hyperglycemic qualities (Hang *et al.*, 2018; Islam *et al.*, 2023). Because of their anti-inflammatory and nephroprotective properties, they are believed to have health benefits. There are number of reported phytochemicals having protective role DN (Table 1).



Figure 1: Different pathways of diabetic nephropathy.

3.1 Alkaloids

3.1.1 Berberine

In an animal model, berberine has proven to be an efficient diabetic nephropathy (DN) treatment. BUN, SCR, proteinuria, KI, IL-6, TNF- α , oxidative stress, and MDA are all indications of improved renal function, as are risk factors TG, TC, and LDL. Because of its antifibrotic, anti-inflammatory, and antioxidative stress capabilities, it has positive benefits (Hu et al., 2022; Ni et al., 2015). By deactivating the TLR4/NF-kB pathways, BBR lowers kidney damage, inflammation, and podocyte death in DN (Zhu et al., 2018). By preventing the growth of the glomerular mesangial matrix through the activation of AMPK, HGSD, a novel berberine solution, enhances kidney function, decreases glomerular volume, activates autophagy, and guards against diabetic kidney disease (Zhang et al., 2020). By suppressing the NLRP3 inflammasome, BBR can reduce tubulointerstitial fibrosis and epithelial-to-mesenchymal transition (EMT) in diabetic kidney disease (DKD) (Ma et al., 2022). Berberine's potential as a DN treatment is suggested by the possibility that it may lessen renal fibrosis, decrease glomerular hypertrophy and mesangial matrix growth, decrease TGF- β and -SMA expression, and lessen kidney damage (Li et al., 2017). Because it controls glucose uptake, inhibits the PI3K/Akt/AS160/GLUT1 signalling pathways, and stops abnormal glomerular mesangial cell (GMC) proliferation, berberine has the potential to treat DN-related renal pathology and glomerular dysfunction (Ni et al., 2022). Berberine has the potential to treat DN by reducing tubulointerstitial fibrosis. It accomplishes this by promoting Nrf2 pathways, blocking TGF-B/Smad/EMT signalling, and lowering oxidative stress, which eventually results

in kidney protection (Zhang *et al.*, 2016). The combination of metformin and berberine displays increased anti-diabetic nephropathy effects by reducing blood sugar, boosting insulin sensitivity, improving lipid metabolism, and lessening renal damage. This combination improves renal function by upregulating Trib1, downregulating C/EBP, inhibiting lipid synthesis proteins, and decreasing NF- κ B signalling (Zhang *et al.*, 2021). Berberine has protective properties in streptozotocin-induced DN *via* modulating the G protein-adenylyl cyclase (AC)-cAMP signalling pathway, lowering glomerular mesangial cell (MC) proliferation, and alleviating renal damage (Tang *et al.*, 2013). Through inhibition of aldose reductase (AR) activity, reduction of oxidative stress, enhancement of extracellular matrix formation, and cell proliferation, berberine exhibits potential in the treatment of renal dysfunction in DN (Liu *et al.*, 2008).

3.1.2 Boldine

Due to its anti-inflammatory, antioxidant, and hypoglycemic properties, the alkaloid boldine from the boldo plant has therapeutic potential. Boldine reduced blood pressure, oxidative stress, elevated blood sugar, and protein leakage in diabetic rats. Additionally, it decreased kidney damage indicators and matrix protein changes (Hernández-Salinas *et al.*, 2013). In hypertension, boldine therapy decreased systolic blood pressure, oxidative stress, and kidney damage. Additionally, it prevented inflammation (ED-1, OPN) and kidney injury (-SMA, COM III) indicators in the 2K1C animals. Boldine stopped RAS mediators (ACE-1, TGF- β) from rising (Gomez *et al.*, 2018).

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3.1.3 Trigonelline

Trigonelline, an alkaloid present in the extract of various plants such as Trigonella foenum-graecum, Coffea sp., Glycine max, and Lycopersicon esculentum, displays diverse biological effects, encompassing cancer treatment, hyperglycemia, high cholesterol, and hormonal imbalances (Mohamadi et al., 2018). Research studies have indicated that trigonelline greatly reduces the expression of FN and collagen IV in mesangial ECM in DKD rats, thereby significantly decreasing oxidative stress and pathological alterations in the kidneys (Shao et al., 2019). In the context of treating human mesangial cells (HMCs) activated by HG, trigonelline was found to dramatically decrease the levels of FN and collagen IV, as well as the hyperproliferation of HMCs. Further, trigonelline prevented the activation of the Wnt/\beta-catenin signaling pathway, thereby decreasing cell cycle progression and apoptosis (Chen et al., 2022). In DKD rats, trigonelline increased the expression of peroxisome proliferator-activated receptor-gamma (PPAR-y) and glucose transporter type 4 (GLUT4) proteins, while suppressing the expression of leptin and tumor necrosis factor alpha (TNF- α) proteins, thus decreasing oxidative stress, inflammation, and kidney cell apoptosis (Li et al., 2019). Trigonelline has been found by (Chen et al., 2021) to upregulate the expression of miR-5189-5p, lower the amount of the inhibitor of hypoxia-inducible factor 1 subunit alpha (HIF1AN), activate the AMPK signaling pathway, increase the level of autophagy, and protect renal mesangial cells (Chen et al., 2021). The administration of trigonelline to mice at a dose of 50 mg/kg for 21 days did not cause any changes in the weight of the liver, kidney, thymus, thyroid, or adrenal gland (Zhou et al., 2012). Additionally, trigonelline has been found to enhance renal function by increasing apoptotic cell death in the kidney, decreasing oxidative stress, and reducing kidney tissue fibrosis and degenerative alterations, which suggests its potential in preventing kidney damage in diabetic rats (Ghule et al., 2012).

3.2 Flavonoids

3.2.1 Quercetin

In DN, quercetin has protective properties. Reduced polyuria, lower blood sugar, and normalised hypertriglyceridemia are the effects of quercetin therapy. Additionally, it lowers proteinuria, lowers creatinine levels, and guards against alterations to the structure of the kidneys (Gomes et al., 2015). The blood glucose level was the only one that quercetin treatment did not improve in diabetic rats with raised kidney weight, urine albumin, serum creatine, blood urea nitrogen, and impaired creatinine clearance. Additionally, TGF-1 and CTGF expressions were elevated in diabetic rats, which quercetin reduced (Lai et al., 2012; Gomes et al., 2014; Elbe et al., 2015). Quercetin dramatically improves renal indicators, lowering creatinine, blood urea nitrogen, urine protein, albumin, malondialdehyde, and interleukin-1 while raising the activity of superoxide dismutase and catalase (Hu et al., 2022; Goswami et al., 2023). The treatment of quercetin considerably reduced oxidative stress (lowered malondialdehyde while raising superoxide dismutase and catalase activity) and enhanced a number of renal function markers. By lowering levels of interleukin-1 (IL-1) and tumour necrosis factor-alpha (TNF- α), it also demonstrated antiinflammatory effects (Feng et al., 2022). Mesangial index, Scr, BUN, proteinuria, BG, and other indicators were decreased by quercetin, whereas HDL-C, SOD, GSH, and other markers were increased. By acting through pathways like PI3K/PKB, AMPK-P38 MAPK, Nrf2/HO-1, etc., quercetin improved kidney disease (Li et al., 2022). Quercetin has potential therapeutic effects in cardiovascular and renal illnesses; however, it is unclear how it affects the growth of glomerular mesangial cells (MC) in early DN. The effect of quercetin on high glucose-induced MC proliferation in mice, which showed that it inhibited MC proliferation through activating the Hippo pathway (Lei et al., 2019). In diabetic situations, quercetin reduces EMT and renal fibrosis, possibly through modifying mTORC1/p70S6K and certain transcription factors (Lu et al., 2015). Early diabetic kidney damage is safely and effectively treated by quercetin, which may do so via enhancing lipid metabolism via the SCAP-SREBP2-LDLr signalling pathway (Jiang et al., 2019). Early diabetic kidney damage is safely and effectively treated by quercetin, which may do so via enhancing lipid metabolism via the SCAP-SREBP2-LDLr signalling pathway (Liu et al., 2022). According to studies, quercetin possesses substantial anti-diabetic properties that could lower blood sugar levels and improve insulin sensitivity. It affects important elements and signalling pathways, including TNF-a, NF-kB, AMPK, AKT, and Nrf2, that are involved in insulin resistance and the onset of type 2 diabetes. By focusing on the underlying mechanisms, quercetin has also shown promise in preventing and treating diabetes complications, including diabetic nephropathy (Yan et al., 2022).

3.2.2 Curcumin

In diabetic rat kidneys and glomerular mesangial cells (GMCs) exposed to high glucose, curcumin lowered SphK1 expression, activity, and S1P synthesis, which in turn reduced FN and TGF-B1 overproduction. AP-1 DNA-binding activity was also decreased by curcumin, and c-Jun-siRNA restored the up-regulation of SphK1 brought on by HG (Huang et al., 2013). The effects of curcumin treatment included decreased albuminuria, mesangial matrix expansion, and renal hypertrophy, as well as decreased collagen IV and fibronectin expression and levels of interleukin-1 β , cleaved caspase-1, and NLRP3 protein. Curcumin may be used to treat diabetic nephropathy by inhibiting the NLRP3 inflammasome (Lu et al., 2017). Curcumin significantly reduced the phosphorylated ERK1/2 and PKC- α and PKC- β 1 activity caused by high hyperglycemia. Additionally, curcumin reduced the expression of ECM proteins such fibronectin and type IV collagen as well as TGF-β1, CTGF, osteopontin, and p300 (Soetikno et al., 2011).

3.2.3 Kaempferol

Kaempferol (3,5,7-trihydroxy-2-[4-hydroxyphenyl]-4H-1benzopyran-4-one) is a natural flavonoids compound. Various sources of this substance can be observed in conventional medicinal practices, including, ginkgo, Sophora japonica and galangal, as well as in consumables such as, cauliflower, gooseberry, cabbage, strawberries, grapes, tomatoes and tea (Burgos-Moron *et al.*, 2011). Kaempferol has anti-inflammatory (Crespo *et al.*, 2008), antioxidative stress (Suh *et al.*, 2009). antitumor (Mylonis *et al.*, 2010), anti-atherosclerotic (Feng *et al.*, 2021), hypoglycemic (Alkhalidy *et al.*, 2018), and hypolipidemic (Torres-Villarreal *et al.*, 2019) effects. Kaempferol treatment prevented weight loss, renal damage, and alterations in biochemical parameters. It also reduced inflammation markers, caspase-3, p38, and increased anti-apoptotic factors. Kaempferol improved glucose, insulin levels, reduced oxidative stress, and upregulated Nrf2/HO-1, suggesting its antioxidant potential in preventing diabetic nephropathy (Alshehri, 2023). A study reported, based on in vitro tests using rat and human renal tubular epithelial cells, indicate that kaempferol inhibits RhoA activation, reduces oxidative stress, pro-inflammatory cytokines, and fibrosis-related markers (Sharma et al., 2019). kaempferol treatment increased GLP-1 and insulin release, improved renal histology, reduced fibrosis, and downregulated key markers of DN (Sharma et al., 2020). The potential of kaempferol as a therapeutic agent to mitigate inflammation. It reduced renal inflammation, fibrosis, and kidney dysfunction in diabetic mice by downregulating TRAF6. In vitro, KPF attenuated high glucoseinduced inflammatory and fibrogenic responses in renal cells by downregulating TRAF6 (Luo et al., 2021). Kaempferol shows protective action against DN by enhancing podocyte autophagy and reducing apoptosis, likely mediated through the AMPK/mTOR pathway (Sheng et al., 2022). Kaempferol effects were associated with increased SIRT1 levels and activity, as well as enhanced acetylation of Nrf2 and NF-kB. The SIRT1 inhibitor, kaempferol alleviates DOX-induced nephropathy by upregulating and activating SIRT1 (Alagal et al., 2022). Kaempferol treatment significantly attenuated kidney injury, and functional disturbances. It reduced oxidative stress, inflammation, and apoptotic markers while promoting antioxidant enzyme levels. Kaempferol's effects were associated with modulation of NF-kB p65 and Nrf2 pathways (Alshehri et al., 2022). Kaempferol attenuated D-ribose-induced changes in mesangial cells, possibly by repairing autophagy and reducing AGE accumulation and ROS production (Zhang et al., 2019).

3.2.4 Rutin

The preventive effect of Rutin on diabetic nephropathy (DN) is closely associated with oxidative stress and various signaling pathways, such as TGF-B1/Smad/ECM and TGF-B1/CTGF/ECM, thus highlighting its potential as a preventive treatment for DN in rats (Hao et al., 2012). Recent studies have revealed that Rutin administration effectively prevents the progression of diabetic nephropathy and cardiomyopathy by improving fibrosis and metabolic acidosis (Ganesan et al., 2020). Furthermore, the combined treatment of Rutin and Selenium significantly reduces the levels of IL-6, NF- κ B, TNF- α , Jak-2, and p-Stat3, indicating a potential renoprotective effect against DN through the upregulation of Nrf-2/ HO-1 and downregulation of Jak-2/Stat3 pathways (Zaghloul et al., 2022). The protective potential of Rutin against diabetic complications is attributed to its ability to decrease the formation of harmful substances associated with hyperglycemia, such as sorbitol, reactive oxygen species, advanced glycation end-products, and inflammatory cytokines (Ghorbani, 2017). Rutin was administered at various doses and found to inhibit high glucose-induced mesangial cell viability, ATP content, and expression of ACTA2 and p38. Furthermore, it improved the cell cycle progression of mesangial cells, thereby highlighting the potential of Rutin as a preventive and therapeutic agent for DN by inhibiting the expression of ACTA2 and p38 (Han et al., 2017). Combination therapy of Rutin and Ramipril successfully restored the antioxidant status, down-regulated endoplasmic reticulum stress markers, reversed gene expression changes, reduced oxidative stress and fibrosis, and lowered side effects (Ganesan et al., 2018). rutin treatment was found to prevent hyperpermeability and dysfunction of the tight junction, and this protective effect was associated with the activation of Nrf2, leading to a decrease in reactive oxygen species and inhibition of the RhoA/ROCK pathway (Wang et

al., 2016). Furthermore, the combination of rutin and Vildagliptin was found to be more effective, leading to improved histological conditions and reduced damage of glomeruli and tubules (Tilethe *et al.*, 2013). Rutin was found to reduce the G0/G1 cell phase percentages, inhibit Smad 2/3, laminin, type IV collagen, and TGF- β 1 mRNA expression, and increase antioxidant capacity, S phase cell percentages, and Smad 7 expression (Tang *et al.*, 2011). Lastly, Rutin effectively improves renal function in 5/6 nephrectomised rats, likely through antioxidation and inhibiting TGF β 1-Smad signaling (Han *et al.*, 2015).

3.2.5 Fisetin

Fisetin, a flavonoid, showed positive outcomes in ameliorating DN. fisetin alleviate podocyte injury and DN by restoring the CDKN1B/ P70S6K pathway, promoting autophagy, and inhibiting NLRP3 inflammasomes (Dong et al., 2022). Fisetin enhanced antioxidative stress in DN through the Nrf2/HO-1/GPX4 pathway, attenuating podocyte injury and DN formation (Qian et al., 2023). fisetin mitigated HFD-induced renal injury by regulating iRhom2/NF-kB and Nrf-2/ HO-1 signaling pathways (Chenxu et al., 2021). Fisetin reduced reactive oxygen species (ROS), advanced glycosylation end products (AGEs), inflammatory cytokines, and extracellular matrix accumulation in the kidney, by inhibiting the TGFB/SMAD signaling pathway and regulating matrix metalloproteinases (MMPs), both in vivo and in vitro fisetin therapeutic effects on kidney fibrosis were mediated by inhibiting CD36 expression (Zou et al., 2023). Fisetin treatment lowered extracellular matrix protein expression, reduced p300 expression, and increased MMP-2 expression (Liu et al., 2014). Luteolin and fisetin on inflammatory responses in human monocytes exposed to high glucose levels. The combination treatment reduced NF-kB activity, cytokine release, and histone acetyltransferase activity, while activating SIRT1 and FOXO3a expressions (Kim et al., 2017).

3.2.6 Silibinin

Silibinin improved the diabetic condition, leading to reduced body weight, HbA1c levels, and serum insulin levels. It prevented kidney injury, reduced oxidative stress, activated the AKT signaling pathway, and decreased the levels of p-GSK-3 β , Bax, and cleaved caspase-3. Overall, silibinin showed potential in ameliorating diabetic nephropathy through the activation of the AKT signaling pathway (Liu et al., 2019; Chu et al., 2018; Islam et al., 2021). Silibinin attenuate renal fibrosis in vitro and in vivo via inhibition of NF-KB (Liu et al., 2019). Diabetic rats exhibited hyperglycemia, hyperlipidaemia, and kidney dysfunction, which were significantly improved with silibinin treatment. It had potential to prevent the progression of early diabetic nephropathy (Jain, 2015). Silibinin significantly increased the antifibrosis effect of valsartan in TGF-B1-treated HK-2 cells via inhibition of TGF-B1 signaling pathway (Liu et al., 2020). The combination of Silibinin and MK-521 notably decreased the expression of fibrosisrelated genes and proteins in cultured kidney cells (HK-2) exposed to TGF-B. In a high-fat diet-induced renal fibrosis mouse model, silibinin enhanced the anti-fibrotic effects of MK-521 (Ma et al., 2020).

3.2.7 Luteolin

Luteolin lowered blood glucose and BUN levels in diabetic rats, raised serum sodium and chloride levels, and hindered glycoprotein deposition and collagen fiber production in kidney. It boosted kidney antioxidant enzymes *via* Nrf2, reduced inflammatory cytokines *via* NF- κ B down-regulation, inhibited PI3K/Akt pathway, and suppressed apoptosis-related proteins (Chen *et al.*, 2023; Mehtab *et al.*, 2021). A study showed luteolin protects against diabetic nephropathy by altering SOD, MDA, and HO-1 levels (Wang *et al.*, 2011). Luteolin has been observed to diminish diabetic nephropathy (DN) primarily through the suppression of inflammatory and oxidative responses. The potential target identified is the STAT3 pathway, which ultimately results in the reduction of renal fibrosis and a delay in the advancement of DN (Zhang *et al.* 2021; Xiong *et al.*, 2020). Luteolin hinder podocyte injury and activation of the NLRP3, making potential to treat diabetic nephropathy (Yu *et al.*, 2019). It restored insulin resistance, dyslipidemia, hyperuricemia, and renal inflammation, as well as inhibited the RIP140/NF- κ B pathway and enhanced insulin signaling (Chen *et al.*, 2016).

3.2.8 Diosmin

Diosmin raised insulin, decreased blood sugar, and increased body weight. It improved the levels of antioxidants (SOD, CAT, GSH, and NO) and normalised the structure of the kidney tissue. Diosmin also supported the maintenance of renal function by reducing NF- κ B levels (Ahmad *et al.*, 2016). When diosmin and crocin were combined, insulin resistance was improved, blood pressure, uric acid, and lipoproteins were decreased, and diabetic nephropathy indicators such albumin excretion rate were alleviated. Additionally, they improved filtration and decreased inflammation and oxidative stress, as shown by decreased serum TNF and inflammatory cells (El-Fawal *et al.*, 2018). Diosmin alleviated the HG-mediated endoplasmic reticulum stress injury in HK-2 *via* restraining the PI3K/AKT pathway (Deng *et al.*, 2022).

3.2.9 Catechin

By blocking AGEs and inflammatory pathways, catechin oral administration may reduce the problems associated with diabetes (Zhu *et al.*, 2014). In mice fed with catechin, circRNA.5549 and circRNA.4712 seem to have favourable effects, especially on the circRNA.5549/miR-29a-5P/Cd36 network. This suggests that a potential treatment strategy for diabetic nephropathy would involve targeting their ceRNA regulatory network (Chen *et al.*, 2022).

3.2.10 Apigenin

Apigenin, a natural flavonoid with anti-inflammatory and antifibrosis properties, has shown promise in DN. Apigenin downregulate inflammatory factors (IL-6, IFN-7, TNF-a), inhibited renal fibrosis markers. apigenin's protective role in DN is mediated through the miR-423-5p-USF2 axis (Hou et al., 2021). Apigenin taken orally decreased body weight, corrected HFD-induced hyperinsulinemia, hyperglycemia, hyperlipidemia, kidney impairment, and collagen deposition. While reducing MDA, TNF- α , IL-6, nuclear NF- κ B p65, and the mRNA levels of TGF- β 1, Bax, and caspase-3, apigenin raised the levels of Nrf2, as well as GSH, SOD, and CAT (Aldayel, 2022). Treatment with apigenin decreased renal dysfunction and oxidative stress. Type IV collagen, fibronectin, and TGF-1 levels decreased, indicating lessened fibrosis. Additionally, it prevented the activation of the mitogen-activated protein kinase (MAPK), reducing inflammation (Cavero and Holzgrefe, 2019; Malik and Arya, 2021). By lowering TGF-1, fibronectin, and type IV collagen, apigenin reduced oxidative stress, fibrosis, and renal failure. Additionally, it inhibited MAPK activation, which reduced inflammation (TNF- α , IL-6, and NF-B expression) and apoptosis (increased Bcl-2, decreased Bax, and raised caspase-3 (Malik *et al.*, 2017; Arya *et al.*, 2018). Both anti-inflammatory (inhibiting the release of inflammatory factors) and antioxidant activity (decreasing the formation of lipid peroxidation) were characteristics of apigenin. Apigenin showed protective effects, and the Nrf2/heme oxygenase-1/NF-B pathway may be responsible for these effects (Zhang *et al.*, 2019).

3.2.11 Genistein

Genistein, a phytoestrogen from soybean, shows potential in treating diabetic complications, as it reduces hyperglycemia, insulin resistance, and offers therapeutic benefits (Jiang et al., 2023; Weng et al., 2019). genistein alleviated diabetic nephropathy by inhibiting the MAPK/NF-kB pathway, enhancing mitochondrial function, and exerting anti-inflammatory effects (Li et al., 2022). It shows promise in inhibiting ECM and TGF-B (Yuan et al., 2009). Genistein influences inflammatory responses and reduces oxidative stress by acting on IL-1, IL-6β, TNF-α, SIRT1, NF-κB, and TGF-β1/smad targets. Improved podocyte autophagy, suppression of RAAS overactivity, and eventual alleviation from diabetic nephropathy result from these actions (Hu et al., 2021). Genistein has the potential to prevent DN through regulating inflammation and oxidative stress, particularly in people with diabetes with medium-to-high blood sugar levels (Kim and Lim, 2013). In addition to increased levels of phospho-tyrosine and phospho-ERK/ERK ratio, diabetic mice displayed heightened levels of oxidative stress and apoptosis. These alterations were reversed by genistein administration, suggesting that its renoprotective impact in diabetes is connected to decreased oxidative stress, inflammatory response, and apoptosis (Elmarakby et al., 2011). By lowering multiple risk variables connected to ovariectomyinduced renal impairment, genistein has the potential to protect kidney function. This entails enhancing insulin resistance, decreasing renal oxidative stress, lowering lipid buildup, and reducing extracellular matrix protein expression (Choi and Song, 2009).

3.2.12 Chrysin

Propolis and mushrooms contain a naturally occurring flavonoid called chrvsin (5.7-dihvdroxvflavone) that has anti-inflammatory. antioxidant, and anticancer activities. Proteinuria, glomerular ultrastructure, and slit diaphragm protein levels were all improved by chrysin treatment. Chrysin's efficiency in reducing diabetesinduced podocyte injury is associated with its focus on ER stress pathways (Kang et al., 2017; Siddiqui et al., 2022). Animal models of chrysin protective properties through controlling oxidative stress, inflammation, and apoptosis (Farkhondeh et al., 2019; Siddiqui et al., 2020). In DN mice, chrysin reduced obesity, insulin resistance, impaired renal function, and pathological damage. Additionally, it reduced inflammation and oxidative stress, both essential for the advancement of DN. Chrysin controlled lipid metabolism by altering downstream proteins including adenosine 52 monophosphateactivated protein kinase (AMPK). A study revealed that lipid metabolic modulation by way of activated AMPK plays a role in how chrysin improves DN (Zhou et al., 2022). Chrysin controlled mesenchymal and epithelial markers, which prevented excessive hyperglycemia from causing EMT. Additionally, it affected the migration and integrity of tubular cells as well as collagen formation. Tight junction proteins were reconstituted by chrysin. This raises the possibility that chrysin can prevent the EMT-driven

tubulointerstitial fibrosis that results from diabetic nephropathy (Kang *et al.*, 2015). Chrysin targets glomerulosclerosis and fibrosis brought on by AGE to reduce the buildup of matrix protein in diabetic glomeruli (Lee *et al.*, 2018).

3.2.13 Hesperidin

Hesperidin improved serum, liver, and kidney parameters and fixed abnormalities in the kidneys. Hesperidin's protective actions in diabetic nephropathy may entail regulation of TGF-1-ILK-Akt signalling. It improved nephropathy-related alterations, restored podocyte function, and lowered TGF-1 signalling (Zhang *et al.*, 2018). Hesperidin dramatically reduced glucose levels, liver and kidney damage indicators, and NF- κ B and SIRT1 levels while controlling oxidative stress and NF- κ B levels (Iskender *et al.*, 2017). Hesperidin treatment dramatically reduced serum and renal tissue FGF-23 levels (Dokumacioglu *et al.*, 2019). Hesperidin decreased oxidative stress and renal dysfunction following I/R damage in diabetic rats, indicating its potential to lessen renal consequences in this situation (Kakadiya *et al.*, 2010).

3.3 Tannins

Resources that are naturally renewable are used to make tannins (Pizzi, 2008; Ramakrishnan and Krishnan, 1994). which phenolic compounds are subsequent to phenol in plants (Hagerman, 2002; Sharma, 2019). Galloyl esters and proanthocyanidins, which can be oligomeric or polymeric, are examples of tannins (Khanbabaee and Van Ree, 2001), generated by plants' secondary metabolism (Lewis and Yamamoto, 1998). produced by biogenetic processes. Tannins have a variety of pharmacological effects, including antioxidant, anti-inflammatory, anticancer, antinutritional, and free radical scavenging properties. They also seem to improve metabolic conditions and postpone the beginning of a number of oxidative stress-related disorders (Smeriglio *et al.*, 2017).

3.3.1 Epigallocatechin gallate

The ability of EGCG to stop apoptosis in rats with diabetic nephropathy brought on by a high-fat diet and STZ. The antiapoptotic properties of EGCG, reduction of TGF, improvement of kidney function, and reduction of diabetic nephropathy (Mohan et al., 2017). Supplementing with EGCG prevented high glucose's effects on enhanced IGF-I and IGF-II production. Additionally, it enhanced arachidonic acid pathways, increased GSH content, and decreased lipid peroxide, which prevented mesangial cell failure (Han and Park, 2007). In diabetic nephropathy, EGCG decreased hyperglycemia, proteinuria, lipid peroxidation, as well as the buildup of advanced glycation end products and associated kidney disease. This suggests that EGCG has the ability to minimise the kidney damage brought on by oxidative stress associated with aberrant glucose metabolism (Yamabe et al., 2006; Yoon et al., 2014). Early-stage DN may benefit from EGCG's ability to activate the Nrf2/ARE pathway by raising nuclear Nrf2 while decreasing nuclear Keap1 (Mohan et al., 2020).

3.3.2 Ellagic acid

Ellagic acid plus metformin may prevent type II diabetes-related nephropathy by regulating blood sugar levels, lowering oxidative stress and inflammation, and combating free radicals. This will eventually result in a reduction in the apoptosis of kidney tissue (Harakeh *et al.*, 2023). Ellagic acid protected against oxidative stress, apoptosis, inflammation, and kidney injury. Reduced NF-κB activity, elevated levels of antioxidant enzymes (GSH, GCL, SOD), and improved Nrf2 nuclear translocation were all associated with this. It also increased Akt and GS3K phosphorylation, decreased Fyn phosphorylation and nuclear accumulation, and inhibited keap1's expression in the cytoplasm and its interaction with Nrf2 (ALTamimi *et al.*, 2021). Through suppression of the HMGB1-TLR4-NF-κB pathway, ellagic acid prevented oxidative renal damage brought on by STZ (Zhou *et al.*, 2019). Ellagic acid decreased oxidative stress and renal impairment. Along with lowering renal pathology and inhibiting NF-κB activation, it also reduced fibronectin and transforming growth factor-beta (TGF-β) expression. Ellagic acid also reduced IL-1β, IL-6, and TNF-α serum levels of proinflammatory cytokines (Ahad *et al.*, 2014).

3.4 Stilbenes

Both food and medicinal plants contain the wide group of secondary plant metabolites known as stilbenes, which are a member of the larger family of natural polyphenols (Su *et al.*, 2022; Khan *et al.*, 2021). The most known and best characterized stilbene is resveratrol (Gambini *et al.*, 2015). Studies on the pharmacological effects of stilbenes also show that they possess a variety of qualities, such as effects that have anticancer, antimicrobial, anti-inflammatory, antioxidant, antidegenerative disorders, antidiabetic, neuroprotective, antiageing, and cardioprotective properties (Teka *et al.*, 2022).

3.4.1 Resveratrol

VEGF and Flk-1 expressions in cultured glomerular cells caused by high glucose were reduced by resveratrol, with effects mediated by Sirt1. Additionally, resveratrol prevented endothelial cells from becoming more permeable and rupturing their cellular junctions when VEGF was present. Resveratrol might lessen DN through controlling angiogenesis (Wen et al., 2013). Resveratrol reduces oxidative stress by restoring normal Mn-SOD activity and glucose-lipid metabolism, which in turn reduces renal damage and increases mitochondrial biogenesis with Mn-SOD failure in the kidney. Resveratrol possesses antioxidative properties through a mechanism independent of AMPK and SIRT1 (Kitada et al., 2011; Li et al., 2021). In patients with DN, resveratrol may be a useful adjuvant to angiotensin receptor blockers (ARBs) for lowering urine albumin excretion (Sattarinezhad et al., 2019). By lowering MDA levels and increasing SOD, CAT, GSH, and GPx activities, resveratrol displays antioxidant benefits. It has the potential to lower the pro-inflammatory cytokine IL-1β (Hu et al., 2022). Resveratrol and Rosuvastatin normalised expression to TGF- 1β , fibronectin, while decreasing antioxidant enzyme activity, raising MDA, and increasing MDA. NF-kB/p65. FoxO1, Nrf2, and Sirt1. Encourage preventing oxidative injury to the kidneys (Hussein and Mahfouz, 2016). Resveratrol prevents diabetes-related damage to renal tissue by obstructing the p38 MAPK/TGF-1β signalling pathway (Qiao et al., 2017). Resveratrol up-regulates AMPK expression and activation, exhibits concurrent pro and anti-inflammatory activities, and protects against oxidative stress, all of which may contribute to its positive effects on the early stage of DN (Chang et al., 2011). Expression of intracellular adhesion molecules-1 and resveratrol PAI-1 in diabetic renal cortex. Additionally, it reduced cell growth by inhibiting the Akt/NF-KB pathways (Xu et al., 2014).

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Phytochemicals		Mechanism of action	References
Alkaloids	Berberine	Improve renal function	Hu et al., 2022
		Reduce inflammation, oxidative	Ni et al., 2015
		Reduces renal injury, inflammation, and podocyte apoptosis in DN by inactivating the TLR4/NF-KB pathways	Zhu et al., 2018
		Alleviate tubulointerstitial fibrosis and attenuate EMT in diabetic kidney disease (DKD) by supressing the NLRP3 inflammasome	Ma et al., 2022
		Activating the Nrf2 pathways, supressing TGF- β /Smad/EMT signalling, and reducing oxidative stress	Zhang et al., 2016
		Ameliorating renal dysfunction in DN by inhibiting aldose reductase	Liu et al., 2008
		Reduces NF-KB signalling, leading to improved renal function	Zhang <i>et al.</i> , 2021
	Boldine	Reduced matrix protein alterations and renal damage markers	Hernández-Salinas et al., 2013
		Decrease systolic blood pressure in hypertension	Gomez and Velarde, 2018
		Inhibited markers of kidney damage (α -SAM, COM III) and inflammation (ED-1, OPN) in the 2K1C rats	
	Trigonelline	Reduced oxidative stress, and increased apoptotic cell death in the	Ghule et al., 2012
		kidney Alleviated degenerative changes and fibrosis in kidney tissue	
		Reduce inflammation, oxidative stress, and kidney cell apoptosis	Li et al., 2019
		Increased peroxisome proliferator-activated receptor-gamma (PPAR γ) and GLUT4 protein expression	
		Significantly alleviated the oxidative stress and pathological changes in the kidneys	Shao <i>et al.</i> , 2019
Flavonoids	Quercetin	Decreases creatinine levels, reduced proteinuria and protected against renal structure changes	Gomes et al., 2015
		Attenuated TGF-β1 and CTGF expressions	Lai <i>et al.</i> , 2012; Gomes <i>et al.</i> , 2014; Elbe <i>et al.</i> , 2015
		Reduced creatinine clearance	
		Improved renal pathology and acts via pathways like PI3K/PKB, AMPK-P38 MAPK, Nrf2/HO-1	Li et al., 2022
		Alleviated oxidative stress	Feng et al., 2022
		Exhibited anti-inflammatory activity by TNF- α and IL-1 β levels	
		Inhibited the EGFR signaling pathway by reducing phosphory- lation of EGFR and ERK1/2	Liu <i>et al.</i> , 2022
	Curcumin	Decreased FN and TGF-B1 overproduction	Huang et al., 2013
		Decreased interleukin-1 β , cleaved caspase-1, and NLRP3 protein levels,	Lu et al., 2017
		Attenuated the expression of TGF-β1, CTGF	Soetikno et al., 2011
	Kaempferol	Reduced inflammation markers, caspase-3, p38, and increased anti-apoptotic factors	Alshehri, 2023
		Reduced renal inflammation, fibrosis, and kidney dysfunction in diabetic mice by downregulating TRAF6	Luo et al., 2021
		Reduced oxidative stress, inflammation, and apoptotic markers while promoting antioxidant enzyme levels	Alshehri et al., 2022
		Reducing AGE accumulation and ROS production	Zhang et al., 2019

Table 1: Different phytochemicals and their protective mechanism of actions in DN

		Reduce apoptosis, likely mediated through the AMPK/mTOR pathway	Sheng et al., 2022
		Improved renal histology, reduced fibrosis, and downregulated key markers of DN	Sharma <i>et al.</i> , 2020.
		Alleviates DOX-induced nephropathy by upregulating and activating SIRT1	Alagal et al., 2022
	Rutin	Decrease in reactive oxygen species and inhibition of the RhoA/ROCK pathway	Wang et al., 2016
		Reduced oxidative stress and fibrosis, down-regulated endo- plasmic reticulum stress markers	Ganesan et al., 2018
		Reduced levels of IL-6, NF- κ B, TNF- α , Jak-2, and p-Stat3	Zaghloul et al., 2022
	Fisetin	Enhanced antioxidative stress in DN mice through the Nrf2/ HO-1/GPX4 pathway	Qian et al., 2023
		Mitigated HFD-induced renal injury by regulating iRhom2/ NF-κB and Nrf-2/HO-1 signaling pathways	Chenxu et al., 2021
		Reduced ROS, AGEs, inflammatory cytokines	Zou et al., 2023
		Inhibiting the TGF-β/SMAD signaling pathway	
		Reduced p300 expression, and increased MMP-2 expression	Liu et al., 2014
		Reduced NF-KB activity	Kim et al., 2017
		activating SIRT1 and FOXO3a expressions	
		Alleviate podocyte injury and DN by restoring the CDKN1B/ P70S6K pathway	Dong et al., 2022
		Enhanced antioxidative stress in DN through the Nrf2/HO-1 /GPX4 pathway	Qian et al., 2023.
	Silibinin	Enhanced the anti-fibrotic effects of MK-521	Ma et al., 2020
		Attenuate renal fibrosis in vitro and in vivo via inhibition of NF- κB	Liu et al., 2019
	Luteolin	Reduced NF-kB levels, supporting renal function maintenance	Ahmad et al., 2016
		Improved insulin resistance, lowered blood pressure, uric acid, and lipoproteins, and mitigated diabetic nephropathy markers	El-Fawal et al., 2018
		Alleviated the HG-mediated endoplasmic reticulum stress injury in HK-2 via restraining the PI3K/AKT pathway	Deng et al., 2022
-	Catechin	Inhibiting AGEs and inflammation pathways	Zhu et al., 2014
	Apigenin	Inhibiting the release of Inflammatory factors	Zhang et al., 2019
		Reducing lipid peroxidation production	
		Reducing TGF- β 1, fibronectin, and type IV collagen	Malik et al., 2001Arya et al., 2018
		Decreased inflammation (TNF-a, IL-6, NF-kB expression	
		Inhibited Mitogen Activated Protein Kinase	Cavero and Holzgrefe, 2019
		Apigenin increased Nrf2 levels, along with GSH, SOD, and CAT,	Aldayel, 2022
		Decreasing MDA, TNF-alpha, IL-6, nuclear NF-κB p65,	
		and mRNA levels of TGF-beta1, Bax, and caspase-3	
	Conistoin	Downregulate inflammatory factors (IL-6, IFN- γ , TNF- α)	Hou et al., 2021
	Genistein	Decreasing expression of extracellular matrix proteins	Elmoraldov at al. 2011
		Improved podocyte autophagy inhibition of PAAS overactivity	Hu et al 2021
		Inhibiting ECM and TGE-R	$\frac{1}{2} \frac{1}{2} \frac{1}$
		Alloviate diabetic nonkronethy by inhibiting the MADWAR of	$L_{i} \text{ at } al = 2002$
		pathway	Li el ül., 2022

	Hesperidin	Suppression of TGF-B1-ILK-Akt signaling	Zhang et al., 2018
		Decreased TGF-B1 signaling	
		Regulating oxidative stress and NF- κ B levels, and decreasing SIRT1 levels	Iskender et al., 2017
		Improved renal dysfunction and oxidative stress post I/R injury	Kakadiya <i>et al.</i> , 2010
	Chrysin	Mitigate matrix protein accumulation in diabetic glomeruli by targeting AGE-associated glomerulosclerosis	Lee et al., 2018
		Improved of DN involves lipid metabolism regulation via activated AMPK	Zhou et al., 2022
Tannins	Epigallocate chin gallate	Activates Nrf2/ARE pathway by reducing Keap1 and increasing nuclear Nrf2	Mohan et al., 2020
		Decreasing advanced glycation end-product accumulation and related kidney pathology,	Yamabe et al., 2006
	Ellagic Acid	Decreased serum levels of pro-inflammatory cytokines (IL-1 β , IL-6, TNF- α)	Ahad et al., 2014
		Inhibited the cytoplasmic expression of keap1 and its inter- action with Nrf2, boosted Akt and GS3K β phosphorylation	AL-Tamimi et al., 2021
Stilbenes	Resveratrol	Reversed increase cell proliferation, through Akt/NF- κ B pathways inhibition	Xu et al., 2014
		Up-regulate AMPK expression and activation	Chang et al., 2011
		Inhibiting the p38 MAPK/TGF β 1 signaling pathway	Qiao et al., 2017
		Decreasing MDA levels and enhancing SOD, CAT, GSH, and GPx activities	Hu et al., 2022
		Reducing IL-1 β , a pro-inflammatory cytokine	

4. Conclusion

In conclusion, diabetic nephropathy represents a complex and devastating complication of diabetes mellitus, driven by a network of intricate molecular pathways. Our exploration of the reninangiotensin-aldosterone system (RAAS), polyol pathway, protein kinase C(PKC) pathway, hexosamine pathway, and advanced glycation end products (AGEs) reveals the multifaceted nature of its pathogenesis. Importantly, the protective potential of phytobio actives, including alkaloids, flavonoids, tannins, and stilbenes, offers a promising avenue for therapeutic intervention. These natural compounds exhibit diverse mechanisms of action, providing hope for the development of novel strategies to combat diabetic nephropathy. Incorporating phytobioactives into treatment regimens holds the potential to enhance patient outcomes and alleviate the burden of this debilitating condition.

Acknowledgements

All the authors are thankful to Integral University for providing excellent in campus research facility.

Conflict of interest

The authors declare no conflicts of interest relevant to this article.

References

Adapa, D. and Sarangi, T. K. (2015). A review on diabetes mellitus: complications, management and treatment modalities. J. Med. Health. Sci., 4(3):1-18.

- Ahad, A.; Ganai, A. A.; Mujeeb, M. and Siddiqui, W. A. (2014). Ellagic acid, an NF-κB inhibitor, ameliorates renal function in experimental diabetic nephropathy. Chemico-Biological Interactions, 219:64-75.
- Ahmed, M. A.; Kishore, G; Khader, H. A. and Kasturirangan, M. N. (2013). Risk factors and management of diabetic nephropathy. Saudi Journal of Kidney Diseases and Transplantation, 24(6):1242-1247.
- Ahmed, S.; Mundhe, N.; Borgohain, M.; Chowdhury, L.; Kwatra, M.; Bolshette, N. and Lahkar, M. (2016). Diosmin modulates the NF-kB signal transduction pathways and downregulation of various oxidative stress markers in alloxan-induced diabetic nephropathy. Inflammation, 39:1783-1797.
- Aldayel, T. S. (2022). Apigenin attenuates high-fat diet-induced nephropathy in rats by hypoglycemic and hypolipidemic effects, and concomitant activation of the Nrf2/antioxidant axis. Journal of Functional Foods, 99:105295.
- Alkhalidy, H.; Moore, W.; Wang, Y.; Luo, J.; McMillan, R. P.; Zhen, W. and Liu, D. (2018). The flavonoid kaempferol ameliorates streptozotocininduced diabetes by suppressing hepatic glucose production. Molecules, 23(9):2338.
- Alshehri, A. S. (2023). Kaempferol attenuates diabetic nephropathy in streptozotocin-induced diabetic rats by a hypoglycaemic effect and concomitant activation of the Nrf-2/Ho-1/antioxidants axis. Archives of Physiology and Biochemistry, 129(4):984-997.
- Alshehri, A. S.; El-Kott, A. F.; El-Kenawy, A. E.; Zaki, M. S. A.; Morsy, K.; Ghanem, R. A. and Abd-Ella, E. M. (2022). The ameliorative effect of kaempferol against C_dCl₂-mediated renal damage entails activation of Nrf2 and inhibition of NF-κB. Environmental Science and Pollution Research, 29(38):57591-57602.

- ALTamimi, J. Z.; AlFaris, N. A.; Alshammari, G. M.; Alagal, R. I.; Aljabryn, D. H.; Aldera, H. and Yahya, M. A. (2021). Ellagic acid protects against diabetic nephropathy in rats by regulating the transcription and activity of Nrf2. Journal of Functional Foods, 79:104397.
- Amir Siddiqui, M.; Badruddeen; Akhta; J.; Uddin, S.; Chandrashekharan, S.M.; Ahmad, M.; Khan, M.I. and Khalid M. (2022). Chrysin modulates protein kinase IKKα/ TBK1, insulin sensitivity and hepatic fatty infiltration in dietind-uced obese mice. Drug Development Research, 83(1):194-207.
- Arya, D. V.; Alik, S. M.; Suchal, K. and Bhatia, J. (2018). A18239 Apigenin ameliorates streptozotocin induced diabetic nephropathy in rats by modulation of oxidative stress, apoptosis and inflammation through MAPK pathway. Journal of Hypertension, 36:e63.
- Bernobich, E.; Cosenzi, A.; Campa, C.; Zennaro, C.; Sasso, F.; Paoletti, S. and Bellini, G. (2004). Antihypertensive treatment and renal damage: amlodipine exerts protective effect through the polyol pathway. Journal of Cardiovascular Pharmacology, 44(3):401-406.
- Burgos-Moron, E.; Perez-Guerrero, C.; Lopez-Lazaro, M. and Calderon-Montano, J. (2011). A review on the dietary flavonoid kaempferol. Mini Rev. Med. Chem., 11(4):298-344.
- Cavero, I. and Holzgrefe, H. H. (2019). 18th world congress of basic and clinical pharmacology: thought-provoking lectures on drug safety issues. Expert Opinion on Drug Safety, 18(12):1145-1148.
- Chang, C. C.; Chang, C. Y.; Wu, Y. T.; Huang, J. P.; Yen, T. H. and Hung, L. M. (2011). Resveratrol retards progression of diabetic nephropathy through modulations of oxidative stress, proinflammatory cytokines, and AMP-activated protein kinase. Journal of Biomedical Science, 18(1):1-10.
- Chen, C.; Shi, Y.; Ma, J.; Chen, Z.; Zhang, M. and Zhao, Y. (2022). Trigonelline reverses high glucose-induced proliferation, fibrosis of mesangial cells via modulation of Wnt signaling pathway. Diabetology and Metabolic Syndrome, 14(1):1-13.
- Chen, C.; Zhu, D.; Zhang, S. and Zhang, W. (2022). Identification of circRNA/ miRNA/mRNA regulatory network involving (+)-catechin ameliorates diabetic nephropathy mice. Food Science and Human Wellness, 11(3):660-668.
- Chen, L. Y.; Cheng, H. L.; Liao, C. K.; Kuan, Y. H.; Liang, T. J.; Tšeng, T. J. and Lin, H. C. (2023). Luteolin improves nephropathy in hyperglycemic rats through antioxidant, anti-inflammatory, and antiapoptotic mechanisms. Journal of Functional Foods, 102:105461.
- Chen, L.; Tian, G.; Tang, W.; Luo, W.; Liu, P. and Ma, Z. (2016). Protective effect of luteolin on streptozotocin-induced diabetic renal damage in mice *via* the regulation of RIP140/NF-κB pathway and insulin signalling pathway. Journal of Functional Foods, 22:93-100.
- Chenxu, G; Xianling, D.; Qin, K.; Linfeng, H.; Yan, S.; Mingxin, X. and Minxuan, X. (2021). Fisetin protects against high fat diet-induced nephropathy by inhibiting inflammation and oxidative stress *via* the blockage of iRhom2/NF-κB signaling. International Immunopharmacology, 92:107353.
- Choi, J. S. and Song, J. (2009). Effect of genistein on insulin resistance, renal lipid metabolism, and antioxidative activities in ovariectomized rats. Nutrition, 25(6):676-685.
- Chu, C.; Li, D.; Zhang, S.; Ikejima, T.; Jia, Y.; Wang, D. and Xu, F. (2018). Role of silibinin in the management of diabetes mellitus and its complications. Archives of Pharmacal Research, 41:785-796.
- Crespo, I.; García-Mediavilla, M. V.; Gutiérrez, B.; Sánchez-Campos, S.; Tuñón, M. J. and González-Gallego, J. (2008). A comparison of the effects of kaempferol and quercetin on cytokine-induced pro-inflammatory status of cultured human endothelial cells. British Journal of Nutrition, 100(5):968-976.

- Deng, J.; Zheng, C.; Hua, Z.; Ci, H.; Wang, G and Chen, L. (2022). Diosmin mitigates high glucose-induced endoplasmic reticulum stress through PI3K/ AKT pathway in HK-2 cells. BMC Complementary Medicine and Therapies, 22(1):1-10.
- Diabetes basics. (2023, June 14). International Diabetes Federation. https://idf.org/about-diabetes/introduction/
- **Dokumacioglu, E.; Iskender, H. and Musmul, A. (2019).** Effect of hesperidin treatment on α-Klotho/FGF-23 pathway in rats with experimentally-induced diabetes. Biomedicine and Pharmacotherapy, **109**:1206-1210.
- Dong, W.; Jia, C.; Li, J.; Zhou, Y.; Luo, Y.; Liu, J. and Chen, Y. (2022). Fisetin attenuates diabetic nephropathy-induced podocyte injury by inhibiting NLRP3 inflammasome. Frontiers in Pharmacology, 13:783706.
- Elbe, H.; Vardi, N. Ý. G.A. R.; Esrefoglu, M. U. K. A. D. D. E. S.; Ates, B.; Yologlu, S. and Taskapan, C. (2015). Amelioration of streptozotocin-induced diabetic nephropathy by melatonin, quercetin, and resveratrol in rats. Human and Experimental Toxicology, 34(1):100-113.
- El-Fawal, R.; El Fayouni, H. M. and Mahmoud, M. F. (2018). Diosmin and crocin alleviate nephropathy in metabolic syndrome rat model: Effect on oxidative stress and low-grade inflammation. Biomedicine and Pharmacotherapy, 102:930-937.
- Elmarakby, A. A.; Ibrahim, A. S.; Faulkner, J.; Mozaffari, M. S.; Liou, G. I. and Abdelsayed, R. (2011). Tyrosine kinase inhibitor, genistein, reduces renal inflammation and injury in streptozotocin-induced diabetic mice. Vascular Pharmacology, 55(5-6):149-156.
- Farkhondeh, T.; Samarghandian, S. and Roshanravan, B. (2019). Impact of chrysin on the molecular mechanisms underlying diabetic complications. Journal of Cellular Physiology, 234(10):17144-17158.
- Feng, X.; Bu, F.; Huang, L.; Xu, W.; Wang, W. and Wu, Q. (2022). Preclinical evidence of the effect of quercetin on diabetic nephropathy: A meta-analysis of animal studies. European Journal of Pharmacology, 921:174868.
- Feng, Z.; Wang, C.; Jin, Y.; Meng, Q.; Wu, J. and Sun, H. (2021). Kaempferolinduced GPER upregulation attenuates atherosclerosis via the PI3K/ AKT/Nrf2 pathway. Pharmaceutical Biology, 59(1):1104-1114.
- Gajjala, P. R.; Sanati, M. and Jankowski, J. (2015). Cellular and molecular mechanisms of chronic kidney disease with diabetes mellitus and cardiovascular diseases as its comorbidities. Frontiers in Immunology, 6:340.
- Gambini, J.; Inglés, M.; Olaso, G; Lopez-Grueso, R.; Bonet-Costa, V.; Gimeno-Mallench, L. and Borras, C. (2015). Properties of resveratrol: *in vitro* and *in vivo* studies about metabolism, bioavailability, and biological effects in animal models and humans. Oxidative Medicine and Cellular Longevity, 2015:837042.
- Ganesan, D.; Albert, A.; Paul, E.; Ananthapadmanabhan, K.; Andiappan, R. and Sadasivam, S. G. (2020). Rutin ameliorates metabolic acidosis and fibrosis in alloxan induced diabetic nephropathy and cardiomyopathy in experimental rats. Molecular and Cellular Biochemistry, 471:41-50.
- Ganesan, D.; Holkar, A.; Albert, A.; Paul, E.; Mariakuttikan, J. and Selvam, G S. (2018). Combination of ramipril and rutin alleviate alloxan induced diabetic nephropathy targeting multiple stress pathways *in vivo*. Biomedicine and Pharmacotherapy, 108:1338-1346.
- Ghaderian, S. B.; Hayati, F.; Shayanpour, S. and Mousavi, S. S. B. (2015). Diabetes and end-stage renal disease; a review article on new concepts. Journal of Renal Injury Prevention, 4(2):28.

- Gheith, O.; Othman, N.; Nampoory, N.; Halimb, M. A.; and Al-Otaibi, T. (2016). Diabetic kidney disease: difference in the prevalence and risk factors worldwide. Journal of The Egyptian Society of Nephrology and Transplantation, 16(3):65.
- Ghorbani, A. (2017). Mechanisms of antidiabetic effects of flavonoid rutin. Biomedicine and Pharmacotherapy, 96:305-312.
- Ghule, A. E.; Jadhav, S. S. and Bodhankar, S. L. (2012). Trigonelline ameliorates diabetic hypertensive nephropathy by suppression of oxidative stress in kidney and reduction in renal cell apoptosis and fibrosis in streptozotocin induced neonatal diabetic (nSTZ) rats. International Immunopharmacology, 14(4):740-748.
- Gomes, I. B.; Porto, M. L.; Santos, M. C.; Campagnaro, B. P.; Gava, A. L.; Meyrelles, S. S. and Vasquez, E. C. (2015). The protective effects of oral low-dose quercetin on diabetic nephropathy in hypercholesterolemic mice. Frontiers in Physiology, 6:247.
- Gomes, L; Porto, M. L.; Santos, M. C. L.; Campagnaro, B. P.; Pereira, T.; Meyrelles, S. S. and Vasquez, E. C. (2014). Renoprotective, anti-oxidative and antiapoptotic effects of oral low-dose quercetin in the C57BL/6J model of diabetic nephropathy. Lipids in Health and Disease, 13(1):1-10.
- Gómez, G. I. and Velarde, V. (2018). Boldine improves kidney damage in the goldblatt 2K1C model avoiding the increase in TGF-β. International Journal of Molecular Sciences, 19(7):1864.
- Goswami, K.; Badruddeen; Arif, M.; Akhtar, J.; Khan, M.I. and Ahmad, M. (2024). Flavonoids, isoflavonoids and others bioactives for insulin sensitizations. Curr Diabetes Rev., 20(2):e270423216247 . https:// dx.doi.org/10.2174/1573399819666230427095200.
- Hagerman, A. E. (2002). The Tannin Handbook, Biological Activity of Tannins. Miami University.
- Han, C. S.; Liu, K.; Zhang, N.; Li, S. W. and Gao, H. C. (2017). Rutin suppresses high glucose-induced ACTA2 and p38 protein expression in diabetic nephropathy. Experimental and Therapeutic Medicine, 14(1):181-186.
- Han, H. J. and Park, S. H. (2007). Epigallocatechin gallate protects the dysfunction of mesangial cells in hyperglycemic Conditions *in vitro*. Laboratory Animal Research, 23(4):447-452.
- Han, Y.; Lu, J. S.; Xu, Y.; Zhang, L. and Hong, B. F. (2015). Rutin ameliorates renal fibrosis and proteinuria in 5/6-nephrectomized rats by antioxidation and inhibiting activation of TGFβ1-smad signaling. International Journal of Clinical and Experimental Pathology, 8(5):4725.
- Haneda, M.; Koya, D.; Isono, M. and Kikkawa, R. (2003). Overview of glucose signaling in mesangial cells in diabetic nephropathy. Journal of the American Society of Nephrology, 14(5):1374-1382.
- Haneda, M.; Koya, D.; Isono, M. and Kikkawa, R. (2003). Overview of glucose signaling in mesangial cells in diabetic nephropathy. Journal of the American Society of Nephrology, 14(5):1374-1382.
- Hao, H. H.; Shao, Z. M.; Tang, D. Q.; Lu, Q.; Chen, X.; Yin, X.X. and Chen, H. (2012). Preventive effects of rutin on the development of experimental diabetic nephropathy in rats. Life sciences, 91(19-20):959-967.
- Harakeh, S.; Saber, S. H.; El-Shitany, N.; Ali, S. S.; Alamri, T.; Al-Rabia, M. W. and Mousa, S. (2023). Mitigation of diabetes type II-induced nephropathy by ellagic acid nanoformulations: Amended glycemic control, oxidative stress, inflammation, and induced apoptosis. Journal of King Saud University-Science, 102774.
- Heilig, C. W; Kreisberg, J. L; Freytag, S.; Murakami, T.; Ebina, Y.; Guo, L. and Brosius III, F. C. (2001). Antisense GLUT-1 protects mesangial cells from glucose induction of GLUT-1 and fibronectin expression. American Journal of Physiology-Renal Physiology, 280(4):F657-F666.

- Hernández-Salinas, R.; Vielma, A. Z.; Arismendi, M. N.; Boric, M. P.; Sáez, J. C. and Velarde, V. (2013). Boldine prevents renal alterations in diabetic rats. Journal of Diabetes Research, 2013:1-13.
- Hou, Y.; Zhang, Y.; Lin, S.; Yu, Y.; Yang, L.; Li, L. and Wang, W. (2021). Protective mechanism of apigenin in diabetic nephropathy is related to its regulation of miR-423-5P-USF2 axis. American Journal of Translational Research, 13(4):2006.
- Hu, H. C.; Lei, Y. H.; Zhang, W. H. and Luo, X. Q. (2022). Antioxidant and antiinflammatory properties of resveratrol in diabetic nephropathy: A systematic review and meta-analysis of animal studies. Frontiers in Pharmacology, 13:841818.
- Hu, Q.; Qu, C.; Xiao, X.; Zhang, W.; Jiang, Y.; Wu, Z. and Zhao, Y. (2021). Flavonoids on diabetic nephropathy: Advances and therapeutic opportunities. Chinese Medicine, 16(1):1-17.
- Hu, S.; Wang, J.; Liu, E.; Zhang, X.; Xiang, J.; Li, W. and Ma, X. (2022). Protective effect of berberine in diabetic nephropathy: A systematic review and meta-analysis revealing the mechanism of action. Pharmacological Research, 185:106481.
- Hu, T.; Yue, J.; Tang, Q.; Cheng, K. W.; Chen, F.; Peng, M. and Wang, M. (2022). The effect of quercetin on diabetic nephropathy (DN): A systematic review and meta-analysis of animal studies. Food and Function, 13(9):4789-4803.
- Huang, J.; Huang, K.; Lan, T.; Xie, X.; Shen, X.; Liu, P. and Huang, H. (2013). Curcumin ameliorates diabetic nephropathy by inhibiting the activation of the SphK1-S1P signaling pathway. Molecular and Cellular Endocrinology, 365(2):231-240.
- Huang, W.; He, J.; Nisar, M. F.; Li, H. and Wan, C. (2018). Phytochemical and pharmacological properties of *Chaenomeles speciosa*: An edible medicinal *Chinese mugua*. Evidence-Based Complementary and Alternative Medicine, 2018:9591845.
- Hussein, M. M. and Mahfouz, M. K. (2016). Effect of resveratrol and rosuvastatin on experimental diabetic nephropathy in rats. Biomedicine and Pharmacotherapy, 82:685-692.
- Iskender, H.; Dokumacioglu, E.; Sen, T. M.; Ince, I.; Kanbay, Y. and Saral, S. (2017). The effect of hesperidin and quercetin on oxidative stress, NF-κB and SIRT1 levels in a STZ-induced experimental diabetes model. Biomedicine and Pharmacotherapy, 90:500-508.
- Islam, A.; Mishra, A.; Ahsan, R. and Fareha, S. (2023). Phytopharmaceuticals and Herbal Approaches to Target Neurodegenerative Disorders. Drug Research, 73(7):388-407. DOI 10.1055/a-2076-7939.
- Islam, A.; Mishra, A.; Siddiqui, M. A. and Siddiquie, S. (2021). Recapitulation of evidence of phytochemical, pharmacokinetic and biomedical application of silybin. Drug Research, 71(09):489-503.
- Jain, M. D. (2015). Silibinin, a bioactive flavanone, prevents the progression of early diabetic nephropathy in experimental type-2 diabetic rats. International Journal of Green Pharmacy, 9(2):118-124.
- Jiang, T.; Dong, Y.; Zhu, W.; Wu, T.; Chen, L.; Cao, Y. and Zhong, T. (2023). Underlying mechanisms and molecular targets of genistein in the management of type 2 diabetes mellitus and related complications. Critical Reviews in Food Science and Nutrition, 3:1-13.
- Jiang, X.; Yu, J.; Wang, X.; Ge, J. and Li, N. (2019). Quercetin improves lipid metabolism via SCAP-SREBP2-LDLr signaling pathway in earlystage diabetic nephropathy. Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy, 12:827-839.
- Kakadiya, J.; Patel, D. and Shah, N. (2010). Effect of hesperidin on renal complication in experimentally induced renal damage in diabetic sprague dawley rats. Journal of Ecobiotechnology, 2(2):45-50.

- Kang, M. K.; Park, S. H.; Choi, Y. J.; Shin, D. and Kang, Y. H. (2015). Chrysin inhibits diabetic renal tubulointerstitial fibrosis through blocking epithelial to mesenchymal transition. Journal of Molecular Medicine, 93:759-772.
- Kang, M. K.; Park, S. H.; Kim, Y. H.; Lee, E. J.; Antika, L. D.; Kim, D. Y. and Kang, Y. H. (2017). Chrysin ameliorates podocyte injury and slit diaphragm protein loss via inhibition of the PERK-eIF2α-ATF-CHOP pathway in diabetic mice. Acta Pharmacologica Sinica, 38(8):1129-1140.
- Khan, M.M.; Badruddeen; Ahmad, U.; Akhtar, J.; Khan, M.I. and Khan, M.F. (2021). Acute and sub-acute oral toxicity of pterostilbene, docosahexaenoic acid and its combination in sprague dawley rats. Biochemical and Cellular Archives, 21(1):1-8.
- Khanbabaee, K. and Van Ree, T. (2001). Tannins: classification and definition. Natural Product Reports, 18(6):641-649.
- Kim, A. and Yun, J. M. (2017). Combination treatments with luteolin and fisetin enhance anti-inflammatory effects in high glucose-treated THP-1 cells through histone acetyltransferase/histone deacetylase regulation. Journal of Medicinal Food, 20(8):782-789.
- Kim, M. J. and Lim, Y. (2013). Protective effect of short-term genistein supplementation on the early stage in diabetes-induced renal damage. Mediators of Inflammation, 2013:510212.
- Kiritoshi, S.; Nishikawa, T.; Sonoda, K.; Kukidome, D.; Senokuchi, T.; Matsuo, T. and Araki, E. (2003). Reactive oxygen species from mitochondria induce cyclooxygenase-2 gene expression in human mesangial cells: potential role in diabetic nephropathy. Diabetes, 52(10):2570-2577.
- Kitada, M., Kume, S., Imaizumi, N., and Koya, D. (2011). Resveratrol improves oxidative stress and protects against diabetic nephropathy through normalization of Mn-SOD dysfunction in AMPK/SIRT1-independent pathway. Diabetes, 60(2):634-643.
- Koche, D.; Shirsat, R. and Kawale, M. A. H. E. S. H. (2016). An overerview of major classes of phytochemicals: their types and role in disease prevention. Hislopia J., 9(1/2):1-11.
- Kolm-Litty, V.; Sauer, U.; Nerlich, A.; Lehmann, R. and Schleicher, E. D. (1998). High glucose-induced transforming growth factor betal production is mediated by the hexosamine pathway in porcine glomerular mesangial cells. The Journal of Clinical Investigation, 101(1):160-169.
- Kumaraswamy, K.P.; Nallaperumal, N.; Karimulla, S.; Devarajan, S.; Jagannathan, P.; Thangavel, V.; Yusuff, I. and Boddu, B. (2022). Formulation and evaluation of capsule of ethanolic extract of Cnidoscolus chayamansa Mc Vaugh leaves for the treatment of diabetes. Ann. Phytomed., 11(2):339-343. http://dx.doi.org/10.54085/ap.2022.11.2.40.
- Lai, P. B.; Zhang, L. and Yang, L. Y. (2012). Quercetin ameliorates diabetic nephropathy by reducing the expressions of transforming growth factor-β1 and connective tissue growth factor in streptozotocininduced diabetic rats. Renal Failure, 34(1):83-87.
- Latha, S. and Vijayakumar, R. (2019). The facts about diabetes mellitus: A review. Galore International Journal of Health Sciences and Research, 4(2):64-75.
- Lee, E. J.; Kang, M. K.; Kim, D. Y.; Kim, Y. H.; Oh, H. and Kang, Y. H. (2018). Chrysin inhibits advanced glycation end products-induced kidney fibrosis in renal mesangial cells and diabetic kidneys. Nutrients, 10(7):882.
- Lei, D.; Chengcheng, L.; Xuan, Q.; Yibing, C.; Lei, W.; Hao, Y. and Qian, L. (2019). Quercetin inhibited mesangial cell proliferation of early diabetic nephropathy through the Hippo pathway. Pharmacological Research, 146:104320.
- Lewis, N. G. and Yamamoto, E. (1989). Tannins-their place in plant metabolism. In: Chemistry and significance of condensed tannins. Boston, MA: Springer US:23-46.

- Li, K. X.; Ji, M. J. and Sun, H. J. (2021). An updated pharmacological insight of resveratrol in the treatment of diabetic nephropathy. Gene, 780:145532.
- Li, Y.; Li, Q.; Wang, C.; Lou, Z. and Li, Q. (2019). Trigonelline reduced diabetic nephropathy and insulin resistance in type 2 diabetic rats through peroxisome proliferator activated receptor γ. Experimental and Therapeutic Medicine, 18(2):1331-1337.
- Li, Y.; Ou, S.; Liu, Q.; Gan, L.; Zhang, L.; Wang, Y. and Wu, W. (2022). Genistein improves mitochondrial function and inflammatory in rats with diabetic nephropathy via inhibiting MAPK/NF-κB pathway. Acta Cirurgica Brasileira, 37(6):e370601.
- Li, Z. and Zhang, W. (2017). Protective effect of berberine on renal fibrosis caused by diabetic nephropathy. Molecular Medicine Reports, 16(2):1055-1062.
- Li, Z; Deng, H.; Guo, X.; Yan, S.; Lu, C.; Zhao, Z and Ma, X. (2022). Effective dose/ duration of natural flavonoid quercetin for treatment of diabetic nephropathy: A systematic review and meta-analysis of rodent data. Phytomedicine, 105:154348.
- Lim, A. K. (2014). Diabetic nephropathy-complications and treatment. International Journal of Nephrology and Renovascular Disease, 7:361-381.
- Liu, K.; Zhou, S.; Liu, J.; Wang, Y.; Zhu, F. and Liu, M. (2019). Silibinin attenuates high-fat diet-induced renal fibrosis of diabetic nephropathy. Drug design, Development and Therapy, 13:3117-3126.
- Liu, R. H. (2013). Health-promoting components of fruits and vegetables in the diet. Advances in Nutrition, 4(3):384S-392S.
- Liu, R.; Wang, Q.; Ding, Z.; Zhang, X.; Li, Y.; Zang, Y. and Zhang, G. (2020). Silibinin augments the antifibrotic effect of valsartan through inactivation of TGF-β1 signaling in kidney. Drug Design, Development and Therapy, 14:603-611.
- Liu, W.; Liu, P.; Tao, S.; Deng, Y.; Li, X.; Lan, T. and Zhou, S. F. (2008). Berberine inhibits aldose reductase and oxidative stress in rat mesangial cells cultured under high glucose. Archives of Biochemistry and Biophysics, 475(2):128-134.
- Liu, Y.; Li, Y.; Xu, L.; Shi, J.; Yu, X.; Wang, X. and Lu, Q. (2022). Quercetin attenuates podocyte apoptosis of diabetic nephropathy through targeting EGFR signaling. Frontiers in Pharmacology, 12:792777.
- Liu, Y.; Ye, J.; Cao, Y.; Zhang, R.; Wang, Y.; Zhang, S. and Ye, S. (2019). Silibinin ameliorates diabetic nephropathy via improving diabetic condition in the mice. European Journal of Pharmacology, 845:24-31.
- Liu, Y.; Zhou, B.; SU, H.; Sun, M. and Shen, J. (2014). Fisetin attenuates diabetic nephropathy by regulating transcriptional coactivator p300 and matrix metalloproteinase-2. Chinese Journal of Endocrinology and Metabolism, 12:146-149.
- Lu, M.; Yin, N.; Liu, W.; Cui, X.; Chen, S. and Wang, E. (2017). Curcumin ameliorates diabetic nephropathy by suppressing NLRP3 inflammasome signaling. BioMed Research International, 2017:1516985.
- Lu, Q.; Ji, X. J.; Zhou, Y. X.; Yao, X. Q.; Liu, Y. Q.; Zhang, F. and Yin, X. X. (2015). Quercetin inhibits the mTORC1/p70S6K signaling-mediated renal tubular epithelial-mesenchymal transition and renal fibrosis in diabetic nephropathy. Pharmacological Research, 99:237-247.
- Luo, W; Chen, X.; Ye, L.; Chen, X.; Jia, W; Zhao, Y. and Wang, Y. (2021). Kaempferol attenuates streptozotocin-induced diabetic nephropathy by downregulating TRAF6 expression: the role of TRAF6 in diabetic nephropathy. Journal of Ethnopharmacology, 268:113553.
- Ma, Z.; Zang, W.; Wang, H. and Wei, X. (2020). Silibinin enhances antirenal fibrosis effect of MK-521 via downregulation of TGF-β signaling pathway. Human Cell, 33:330-336.

- Ma, Z; Zhu, L; Wang, S.; Guo, X.; Sun, B.; Wang, Q. and Chen, L. (2022). Berberine protects diabetic nephropathy by suppressing epithelial-tomesenchymal transition involving the inactivation of the NLRP3 inflammasome. Renal Failure, 44(1):923-932.
- Malik, S. and Arya, D. S. (2021). Apigenin attenuates diabetic nephropathy in Type 1 diabetic rats. Journal of Hypertension, 39:e210.
- Malik, S.; Suchal, K.; Khan, S. I.; Bhatia, J.; Kishore, K.; Dinda, A. K. and Arya, D. S. (2017). Apigenin ameliorates streptozotocin-induced diabetic nephropathy in rats *via* MAPK-NF-κB-TNF-α and TGF-β1-MAPK-fibronectin pathways. American Journal of Physiology-Renal Physiology, 313(2):F414-F422.
- Mehta, C.; Kushwaha, K. and Gupta, J. (2021). Luteolin: protective effects against diabetes and diabetes associated complications. Romanian Journal of Diabetes Nutrition and Metabolic Diseases, 28(3):303-310.
- Menne, J.; Park, J. K.; Boehne, M.; Elger, M.; Lindschau, C.; Kirsch, T. and Haller, H. (2004). Diminished loss of proteoglycans and lack of albuminuria in protein kinase C-α-Deficient diabetic mice. Diabetes, 53(8):2101-2109.
- Mohamadi, N.; Sharififar, F.; Pournamdari, M. and Ansari, M. (2018). A review on biosynthesis, analytical techniques, and pharmacological activities of trigonelline as a plant alkaloid. Journal of Dietary Supplements, 15(2):207-222.
- Mohan, T.; Narasimhan, K. K. S.; Ravi, D. B.; Velusamy, P.; Chandrasekar, N.; Chakrapani, L. N. and Periandavan, K. (2020). Role of Nrf2 dysfunction in the pathogenesis of diabetic nephropathy: Therapeutic prospect of epigallocatechin-3-gallate. Free Radical Biology and Medicine, 160:227-238.
- Mohan, T.; Velusamy, P.; Chakrapani, L. N.; Srinivasan, A. K.; Singh, A.; Johnson, T. and Periandavan, K. (2017). Impact of EGCG supplementation on the progression of diabetic nephropathy in rats: An insight into fibrosis and apoptosis. Journal of Agricultural and Food Chemistry, 65(36):8028-8036.
- Mylonis, I.; Lakka, A.; Tsakalof, A. and Simos, G. (2010). The dietary flavonoid kaempferol effectively inhibits HIF-1 activity and hepatoma cancer cell viability under hypoxic conditions. Biochemical and Biophysical Research Communications, 398(1):74-78.
- Ni, W. J.; Ding, H. H. and Tang, L. Q. (2015). Berberine as a promising antidiabetic nephropathy drug: An analysis of its effects and mechanisms. European Journal of Pharmacology, 760:103-112.
- Ni, W. J.; Guan, X. M.; Zeng, J.; Zhou, H.; Meng, X. M. and Tang, L. Q. (2022). Berberine regulates mesangial cell proliferation and cell cycle to attenuate diabetic nephropathy through the PI3K/Akt/AS160/GLUT1 signalling pathway. Journal of Cellular and Molecular Medicine, 26(4):1144-1155.
- Nishikawa, T.; Kukidome, D.; Sonoda, K.; Fujisawa, K.; Matsuhisa, T.; Motoshima, H. and Araki, E. (2007). Impact of mitochondrial ROS production on diabetic vascular complications. Diabetes Research and Clinical Practice, 77(3):S41-S45.
- Okur, M. E.; Karantas, I. D. and Siafaka, P. I. (2017). Diabetes mellitus: A review on pathophysiology, current status of oral medications and future perspectives. Acta Pharm. Sci., 55(1):61-82.
- Pizzi, A. (2008). Tannins: Major sources, properties and applications. In Monomers, polymers and composites from renewable resources. Elsevier, pp:179-199.
- Punthakee, Z.; Goldenberg, R.; and Katz, P. (2018). Definition, classification and diagnosis of diabetes, prediabetes and metabolic syndrome. Canadian Journal of Diabetes, 42:S10-S15.

- Qian, X.; Lin, S.; Li, J.; Jia, C.; Luo, Y.; Fan, R. and Chen, Y. (2023). Fisetin ameliorates diabetic nephropathy-induced podocyte injury by modulating Nrf2/HO-1/GPX4 signaling pathway. Evidence-Based Complementary and Alternative Medicine, 23:210.
- Qiao, Y.; Gao, K.; Wang, Y.; Wang, X. and Cui, B. O. (2017). Resveratrol ameliorates diabetic nephropathy in rats through negative regulation of the p38 MAPK/TGF-β1 pathway. Experimental and Therapeutic Medicine, 13(6):3223-3230.
- Rahimi, Z. (2016). The role of renin angiotensin aldosterone system genes in diabetic nephropathy. Canadian Journal of Diabetes, 40(2)1:78-183.
- Ramakrishnan, K. and Krishnan, M. R. V. (1994). Tannin-classification, analysis and applications. Ancient Science of Life, 13(3-4):232.
- Sato, A.; Hayashi, K.; Naruse, M. and Saruta, T. (2003). Effectiveness of aldosterone blockade in patients with diabetic nephropathy. Hypertension, 41(1):64-68.
- Sattarinezhad, A.; Roozbeh, J.; Yeganeh, B. S.; Omrani, G. R. and Shams, M. (2019). Resveratrol reduces albuminuria in diabetic nephropathy: A randomized double-blind placebo-controlled clinical trial. Diabetes and Metabolism, 45(1):53-59.
- Shao, X.; Chen, C.; Miao, C.; Yu, X.; Li, X.; Geng, J. and Shi, Y. (2019). Expression analysis of microRNAs and their target genes during experimental diabetic renal lesions in rats administered with ginsenoside Rb1 and trigonelline. Die Pharmazie. An International Journal of Pharmaceutical Sciences, 74(8):492-498.
- Sharma, D.; Gondaliya, P.; Tiwari, V. and Kalia, K. (2019). Kaempferol attenuates diabetic nephropathy by inhibiting RhoA/Rho-kinase mediated inflammatory signalling. Biomedicine and Pharmacotherapy, 109:1610-1619.
- Sharma, D.; Tekade, R. K. and Kalia, K. (2020). Kaempferol in ameliorating diabetes-induced fibrosis and renal damage: An *in vitro* and *in vivo* study in diabetic nephropathy mice model. Phytomedicine, 76:153235.
- Sharma, K. P. (2019). Tannin degradation by phytopathogen's tannase: A Plant's defense perspective. Biocatalysis and Agricultural Biotechnology, 21:101342.
- Siddiqui, M.A.; Badruddeen.; Akhtar, J.; Uddin, M.S. S.; Irfan Khan, M. and Khalid, M. (2020). Molecular mechanism of interactions between chrysin and I-kappa-B kinase epsilon (IKKe)/tank binding kinase-1 (TBK1): Cell-based assay and *in silico* molecular docking studies. Journal of Biomolecular Structure and Dynamics, 38(2):589-96.
- Silva, L. C. L.; de Souza, G. H.; Pateis, V. D. O.; Ames-Sibin, A. P.; Silva, B. P.; Bracht, L. and Sá-Nakanishi, A. B. (2023). Inhibition of gluconeogenesis by boldine in the perfused liver: Therapeutical implication for glycemic control. International Journal of Hepatology, 2023:1283716.
- Singh, A. K.; Mo, W. A.; Dunea, G. and Arruda, J. A. (1998). Effect of glycated proteins on the matrix of glomerular epithelial cells. Journal of the American Society of Nephrology, 9(5):802-810.
- Smeriglio, A.; Barreca, D.; Bellocco, E. and Trombetta, D. (2017). Proanthocyanidins and hydrolysable tannins: Occurrence, dietary intake and pharmacological effects. British Journal of Pharmacology, 174(11): 1244-1262.
- Soetikno, V.; Watanabe, K.; Sari, F. R.; Harima, M.; Thandavarayan, R. A.; Veeraveedu, P. T. and Suzuki, K. (2011). Curcumin attenuates diabetic nephropathy by inhibiting PKCα and PKCβ1 activity in streptozotocin induced type I diabetic rats. Molecular Nutrition and Food Research, 55(11):1655-1665.

- Su, X.; Zhou, D. and Li, N. (2022). Bioactive stilbenes from plants. Studies in Natural Products Chemistry, 73:265-403.
- Suh, K. S.; Choi, E. M.; Kwon, M.; Chon, S.; Oh, S.; Woo, J. T. and Kim, Y. S. (2009). Kaempferol attenuates 2-deoxy-d-ribose-induced oxidative cell damage in MC3T3-E1 osteoblastic cells. Biological and Pharmaceutical Bulletin, 32(4):746-749.
- Suzuki, D.; Miyata, T.; Saotome, N.; Horie, K.; Inagi, R.; Yasuda, Y. and Kurokawa, K. (1999). Immunohistochemical evidence for an increased oxidative stress and carbonyl modification of proteins in diabetic glomerular lesions. Journal of the American Society of Nephrology, 10(4):822-832.
- Tang, D. Q.; Wei, Y. Q.; Gao, Y. Y.; Yin, X. X.; Yang, D. Z.; Mou, J. and Jiang, X. L. (2011). Retracted: protective effects of rutin on rat glomerular mesangial cells cultured in high glucose conditions. Phytotherapy Research, 25(11):1640-1647.
- Tang, L. Q.; Wang, F. L.; Zhu, L. N.; Lv, F.; Liu, S. and Zhang, S. T. (2013). Berberine ameliorates renal injury by regulating G proteins-AC-cAMP signaling in diabetic rats with nephropathy. Molecular Biology Reports, 40:3913-3923.
- Teka, T.; Zhang, L.; Ge, X.; Li, Y.; Han, L. and Yan, X. (2022). Stilbenes: Source plants, chemistry, biosynthesis, pharmacology, application and problems related to their clinical Application-A comprehensive review. Phytochemistry, 197:113128.
- Tilethe, S.; Chourasiya, P. K.; Dhakad, R. S. and Kumar, D. (2013). Potential of rutin and vildagliptin combination against alloxan induced diabetic nephropathy in mice. Research Journal of Pharmaceutical Sciences, ISSN 2319, 555X.
- Torres-Villarreal, D.; Camacho, A.; Castro, H.; Ortiz-Lopez, R. and De la Garza, A. L. (2019). Antiobesity effects of kaempferol by inhibiting adipogenesis and increasing lipolysis in 3T3-L1 cells. Journal of Physiology and Biochemistry, 75:83-88.
- Vasavada, N. and Agarwal, R. (2005). Role of oxidative stress in diabetic nephropathy. Advances in Chronic Kidney Disease, 12(2):146-154.
- Wang, G G; Lu, X. H.; Li, W.; Zhao, X. and Zhang, C. (2011). Protective effects of luteolin on diabetic nephropathy in STZ-induced diabetic rats. Evidence-based Complementary and Alternative Medicine, 2011:323171.
- Wang, X.; Zhao, X.; Feng, T.; Jin, G and Li, Z (2016). Rutin prevents high glucoseinduced renal glomerular endothelial hyperpermeability by inhibiting the ROS/Rhoa/ROCK signaling pathway. Planta Medica, 82(14):1252-1257.
- Weigert, C.; Brodbeck, K.; Sawadogo, M.; Haring, H. U. and Schleicher, E. D. (2004). Upstream stimulatory factor (USF) proteins induce human TGFβ1 gene activation via the glucose-response element–1013/–1002 in mesangial cells: Up-regulation of usf activity by the hexosamine biosynthetic pathway. Journal of Biological Chemistry, 279(16): 15908-15915.
- Wen, D.; Huang, X.; Zhang, M.; Zhang, L.; Chen, J.; Gu, Y. and Hao, C. M. (2013). Resveratrol attenuates diabetic nephropathy *via* modulating angiogenesis. PloS One, 8(12):e82336.
- Weng, L.; Zhang, F.; Wang, R.; Ma, W. and Song, Y. (2019). A review on protective role of genistein against oxidative stress in diabetes and related complications. Chemico-Biological Interactions, 310:108665.
- Xiong, C.; Wu, Q.; Fang, M.; Li, H.; Chen, B. and Chi, T. (2020). Protective effects of luteolin on nephrotoxicity induced by long-term hyperglycaemia in rats. Journal of International Medical Research, 48(4):030006 0520903642.

- Xu, F; Wang, Y; Cui, W; Yuan, H.; Sun, J.; Wu, M. and Miao, L. (2014). Resveratrol prevention of diabetic nephropathy is associated with the suppression of renal inflammation and mesangial cell proliferation: Possible roles of Akt/NF-κB pathway. International Journal of Endocrinology, 2014:289327.
- Yamabe, N.; Yokozawa, T.; Oya, T. and Kim, M. (2006). Therapeutic potential of (-)-epigallocatechin 3-O-gallate on renal damage in diabetic nephropathy model rats. Journal of Pharmacology and Experimental Therapeutics, 319(1):228-236.
- Yan, L.; Vaghari-Tabari, M.; Malakoti, F.; Moein, S.; Qujeq, D.; Yousefi, B. and Asemi, Z. (2022). Quercetin: An effective polyphenol in alleviating diabetes and diabetic complications. Critical Reviews in Food Science and Nutrition, 2022:1-24.
- Yoon, S. P.; Maeng, Y. H.; Hong, R.; Lee, B. R.; Kim, C. G; Kim, H. L. and Shin, B. C. (2014). Protective effects of epigallocatechin gallate (EGCG) on streptozotocin-induced diabetic nephropathy in mice. Acta Histochemical, 116(8):1210-1215.
- Yu, Q.; Zhang, M.; Qian, L.; Wen, D. and Wu, G. (2019). Luteolin attenuates high glucose-induced podocyte injury via suppressing NLRP3 inflammasome pathway. Life Sciences, 225:1-7.
- Yuan, W. J.; Jia, F. Y. and Meng, J. Z. (2009). Effects of genistein on secretion of extracellular matrix components and transforming growth factor beta in high-glucose-cultured rat mesangial cells. Journal of Artificial Organs, 12:242-246.
- Zaghloul, R. A.; Abdelghany, A. M. and Samra, Y. A. (2022). Rutin and selenium nanoparticles protected against STZ-induced diabetic nephropathy in rats through downregulating Jak-2/Stat3 pathway and upregulating Nrf-2/HO-1 pathway. European Journal of Pharmacology, 933:175289.
- Zhang, B.; Zhang, X.; Zhang, C.; Sun, G. and Sun, X. (2021). Berberine improves the protective effects of metformin on diabetic nephropathy in db/db mice through Trib1-dependent inhibiting inflammation. Pharmaceutical Research, 38:1807-1820.
- Zhang, J.; Zhao, X.; Zhu, H.; Wang, J.; Ma, J. and Gu, M. (2019). Apigenin protects against renal tubular epithelial cell injury and oxidative stress by high glucose via regulation of NF-E2-related factor 2 (Nrf2) pathway. Medical Science Monitor: International Medical Journal of Experimental and Clinical Research, 25:5280.
- Zhang, M.; He, L.; Liu, J. and Zhou, L. (2021). Luteolin attenuates diabetic nephropathy through suppressing inflammatory response and oxidative stress by inhibiting STAT3 pathway. Experimental and Clinical Endocrinology and Diabetes, 129(10):729-739.
- Zhang, M.; Zhang, Y.; Xiao, D.; Zhang, J.; Wang, X.; Guan, F. and Chen, L. (2020). Highly bioavailable berberine formulation ameliorates diabetic nephropathy through the inhibition of glomerular mesangial matrix expansion and the activation of autophagy. European Journal of Pharmacology, 873:172955.
- Zhang, N.; Zhao, S.; Hong, J.; Li, W. and Wang, X. (2019). Protective effects of kaempferol on D-ribose-induced mesangial cell injury. Oxidative Medicine and Cellular Longevity, 2019:7564207.
- Zhang, X.; He, H.; Liang, D.; Jiang, Y.; Liang, W.; Chi, Z. H. and Ma, J. (2016). Protective effects of berberine on renal injury in streptozotocin (STZ)-induced diabetic mice. International Journal of Molecular Sciences, 17(8):1327.
- Zhang, Y.; Wang, B.; Guo, F.; Li, Z. and Qin, G. (2018). Involvement of the TGFβ1-ILK-Akt signaling pathway in the effects of hesperidin in type 2 diabetic nephropathy. Biomedicine and Pharmacotherapy, 105:766-772.

- Zhou, B.; Li, Q.; Wang, J.; Chen, P. and Jiang, S. (2019). Ellagic acid attenuates streptozocin induced diabetic nephropathy via the regulation of oxidative stress and inflammatory signaling. Food and Chemical Toxicology, 123:16-27.
- Zhou, Y.; Tao, H.; Xu, N.; Zhou, S.; Peng, Y.; Zhu, J. and Chang, Y. (2022). Chrysin improves diabetic nephropathy by regulating the AMPK mediated lipid metabolism in HFD/STZ induced DN mice. Journal of Food Biochemistry, 46(12):e14379.
- Zhu, D.; Wang, L.; Zhou, Q.; Yan, S.; Li, Z.; Sheng, J. and Zhang, W. (2014). (+) Catechin ameliorates diabetic nephropathy by trapping methylgly-

oxal in type 2 diabetic mice. Molecular Nutrition and Food Research, **58**(12):2249-2260.

- Zhu, L.; Han, J.; Yuan, R.; Xue, L. and Pang, W. (2018). Berberine ameliorates diabetic nephropathy by inhibiting TLR4/NF-κB pathway. Biological Research, 51:1-12.
- Zou, T. F.; Liu, Z. G.; Cao, P. C.; Zheng, S. H.; Guo, W.T.; Wang, T. X. and Yang, X. X. (2023). Fisetin treatment alleviates kidney injury in mice with diabetes-exacerbated atherosclerosis through inhibiting CD36/ fibrosis pathway. Acta Pharmacological Sinica, 20:1-10.

Mohd Hashim, Badruddeen, Juber Akhtar, Mohammad Irfan Khan, Mohammad Ahmad, Anas Islam, Asad Ahmad citation and Fatima Zahra (2023). Diabetic nephropathy: An outline on molecular mechanism and protective pathways of phytoconstituents. J. Phytonanotech. Pharmaceut. Sci., 3(3):1-16. http://dx.doi.org/10.54085/jpps.2023.3.3.1